106. Pterin Chemistry

Part 891)

Natural Primapterin Belongs to the L-Series of 7-(Polyhydroxypropyl)pterins²)

by Max Viscontini

Organisch-chemisches Institut der Universität, Winterthurerstrasse 190, CH-8057 Zürich and Abteilung Klinische Chemie, Kinderspital der Universität, Steinwiesstrasse 75, CH-8032 Zürich

(4.IV.90)

Natural primapterin (80 µg) was isolated from 200 ml of urine of a patient with mild hyperphenylalaninemia. The CD curve of the compound showed that it belongs to the L-series of 7-(polyhydroxypropyl)pterins.

Primapterin has been isolated from the urine of a patient with mild hyperphenylalaninemia [3] and has been found in two other children with the same symptoms [4]. HPLC comparison with reference products showed that primapterin coeluated with 7-(Lerythro-1',2'-dihydroxypropyl)pterin (1) already synthesized by Green and Rembold [5]. This structure, without stereochemical identification, has been confirmed by GC-MS [6].

A 7-(1',2'-dihydroxypropyl)pterin structure may occur in four different diastereoisomeric forms, *i.e.* 1-4, the presence in the urine of two of which, namely 3 and 4, were excluded by HPLC. But neither HPLC nor GC-MS can distinguish if the *erythro*-configuration of primapterin belongs to the D-*erythro* (2) or the L-*erythro* (1) series. To answer this question, we isolated *ca.* 80 µg of natural primapterin, and almost the same quantity of biopterin for comparison, from 200 ml of urine of a patient. CD curves of these compounds compared with those of synthetic substances showed that both belong to the L-series.

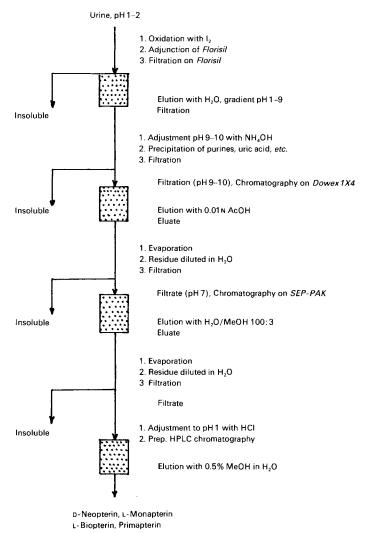
Experimental Part

1. General. Purification of the pterins from urine (Scheme) according to the outlines given in [3]; Florisil and Dowex 1×4 , Fluka AG (CH-9470 Buchs); SEP-PAK C_{18} cartridges; anal. HPLC according to [7]; prep. HPLC according to Matasovic [8]; TLC: silica gel, Merck 60 F_{254} ; eluant: i-PrOH/3% boric acid soln. 4:1. CD: Jasco-J-500A instrument [9].

Presented at the 4th International Conference of Pteridines and Related Biogenic Amines, St. Moritz, March 3-8, 1990. Part 88: [1].

²⁾ We use in this publication the designation rules for pterin derivatives proposed in [2].





2. Purification of Pterins. In a beaker with magnetic stirrer, the pH of a patient's urine was adjusted to 2–3 with 1N HCl and oxidized with a 7% I₂ soln. in MeOH, avoiding an excess of I₂ (KI/starch-paper). Then Florisil was slowly added by stirring (r.t., 1 h) until the urine soln. was nearly colorless and showed a strong blue fluorescence. It was then divided into two portions of 100 ml, and each portion was applied to a Florisil column (13 × 30 mm) and washed with 0.1N HCl and H₂O. The pterins were eluated with the following soln.: 0.05N HCl, H₂O, 0.01N NH₄OH (pterin test with TLC). The slightly yellow soln. was brought to pH 9–10 with 2N NH₄OH. Purines and uric acid precipitated and were filtered off. The basic filtrate was applied to a Dowex 1 × 4 column (26 × 40 mm) and washed with H₂O to neutrality. The filtrate must be free of pterins (TLC). Elution occurred with 0.01N AcOH. During this operation, pterins were separated from most salts, purines, uric acid, and other components of the urine. HPLC showed in the AcOH eluate D-neopterin, primapterin, and L-biopterin in almost equal concentration, a small quantity of L-monapterin, and traces of pterin. The compounds were not pure enough for prep. separation. Therefore, the soln. was evaporated once more. Then 4–5 ml H₂O were added to the residue and the insoluble parts

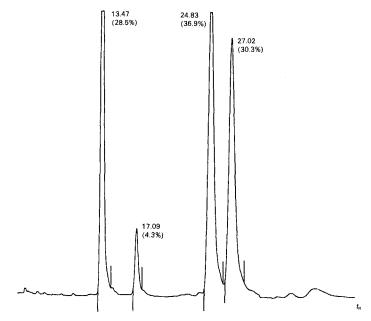


Fig. 1. Anal. HPLC of the pterin solution ready for prep. HPLC. 0.5% MeOH/H2O.

filtered off. Fractions of 1 ml of the filtrate were purified by chromatography on SEP-PAK cartridges which were washed with $H_2O/MeCN$ 4:1 (25 ml). Elution was carried out with 25 ml of $H_2O/MeOH$ 100:3, and then 5 ml of $H_2O/MeOH/i$ -PrOH 90:5:5. The collected eluates were evaporated, diluted in 7–8 ml H_2O , centrifuged for final elimination of the insoluble parts, adjusted to pH 1–2 with HCl and analyzed for pterins by prep. HPLC (Fig. 1): D-neopterin (28.5%), L-monapterin (4.3%), L-biopterin (36.9%), and primapterin (30.3%).

3. Separation of the Single Pterins and Configuration of Primapterin. The separation of the pterins was performed with fractions of 0.5 ml of liquid, applied to a prep. HPLC column N°SP ODS 05 83 11 074

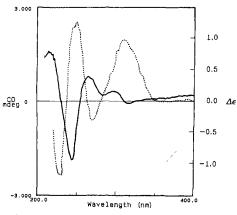


Fig. 2. CD spectrum of L-biopterin isolated from urine (-; ca. 80 µg in 4.5 ml of 0.1N HCl) and mirror image of CD spectrum of synthetic L-biopterin (···)

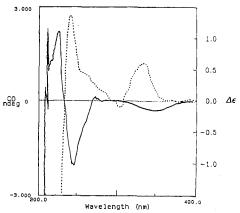


Fig. 3. CD spectrum of primapterin isolated from urine $(-; ca. 80 \mu g \text{ in 4.5 ml of 0.1n HCl})$ and mirror image of CD spectrum of synthetic L-primapterin $(\cdot \cdot \cdot)$

 $(20.5 \times 250 \text{ mm}; Bischoff Analysentechnik, CH-7250 \text{ Leonberg})$, with adsorbent Spherisorb ODS, 5μ ; eluant 0.5 % MeOH, isocratic, flow 9 ml/min. After separation, the eluates of L-biopterin and primapterin were reduced i.v. to 4.5 ml, adjusted to pH 1, and the CD curves were measured and compared with the mirror image of the known curves of synthetic L-biopterin and L-primapterin [9]. The comparison showed distinctly that both isolated pterins belong to the L-series (Figs. 2 and 3).

I am grateful to Prof. Dr. H.-Ch. Curtius and Dr. C. W. Heizmann who received me as host in their laboratory, to Mrs. A. Matasovic, Mrs. C. Adler, and Mr. F. Neuheiser for discussions and technical assistance, Mrs. L. Kierat for anal. HPLC and Mr. P. Uebelhart for the measurement of CD curves. This work was supported by the Swiss National Science Foundation (project No. 31–9427.88).

REFERENCES

- [1] C. Adler, H.-Ch. Curtius, S. Datta, M. Viscontini, Helv. Chim. Acta 1990, 73, 1058.
- [2] M. Viscontini, in 'Pteridines and Biogenic Amines in Neuropsychiatry, Pediatrics, and Immunology', Eds. R. A. Levine, S. Milstien, D. M. Kuhn, and H.-Ch. Curtius, Lakeshore Publishing Comp., Grosse Pointe, Michigan (USA), 1989, pp. 33-50.
- [3] H.-Ch. Curtius, T. Kuster, A. Matasovic, N. Blau, J.-L. Dhondt, Biochem. Biophys. Res. Commun. 1988, 153, 715.
- [4] N. Blau, H.-Ch. Curtius, T. Kuster, A. Matasovic, G. Schoedon, J. L. Dhondt, P. Guibaud, T. Giudici, M. Blaskovics, J. Inher. Metab. Dis., 12. Suppl. 1989, 2, 335.
- [5] B. Green, H. Rembold, Chem. Ber. 1966, 99, 2162.
- [6] T. Kuster, A. Matasovic, N. Blau, J.-L. Dhondt, P. Guibaud, H.-Ch. Curtius, in 'Pteridines and Biogenic Amines in Neuropsychiatry, Pediatrics, and Immunology', Eds. R. A. Levine, S. Milstien, D. M. Kuhn, and H.-Ch. Curtius, Lakeshore Publishing Comp., Grosse Pointe, Michigan (USA), 1989, pp. 83–92.
- [7] A. Niederwieser, W. Staudenmann, E. Wetzel, in 'Biochemistry and Clinical Aspects of Pteridines', Eds. H. Wachter, H.-Ch. Curtius, and W. Pfleiderer, Walter de Gruyter, Berlin-New York, 1982, Vol. 1, pp. 81-102.
- [8] A. Matasovic, personal communication (Abteilung Klinische Chemie, Kinderspital der Universität, CH-8032 Zürich).
- [9] M. Viscontini, R. Bosshard, Helv. Chim. Acta 1990, 73, 337.